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Assessment the *Promachus leoninus* Loew 1848 (Diptera: Asilidae) Species, with COI and NADH₂ gene regions, with new locality records in Anatolia

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Abstract

Promachus leoninus Loew 1848 is known to be distributed just in the western part of Anatolia for a very long time. In the present study additional information is given about the distribution of this species with a new and previous localities map. Assessment of new populations evaluated phylogenetically with the COI and NADH₂ gene regions. As a result of this analysis the new populations are grouped into 3 different diverging populations. Genital features and habitus of the male specimen are photographed.

Keywords: Asilidae, phylogeny, new locality, COI, NADH2 Turkey

Promachus leoninus Loew 1848 (Diptera: Asilidae) Türünün COI ve NADH2 gen bölgeleri ile değerlendirilmesi ve Anadolu'da yeni lokalite kayıtları

Özet

Promachus leoninus Loew 1848'un çok uzun zamandan beri Anadolu'nun sadece batısında yayılış gösterdiği bilinmektedir. Bu çalışmada, yeni ve eski yerleşim haritası ile bu türün yayılışları hakkında ek bilgi verilmektedir. Elde edilen yeni popülasyonlar COI ve NADH₂ gen bölgeleri ile birlikte filogenetik olarak değerlendirilmiştir. Analizler sonucunda yeni populasyonların birbirinden farklılaşmakta olan 3 populasyon şeklinde gruplandığı görülmüştür. Erkek bireyin genital özellikleri ve habitusu fotoğraflanmıştır.

Anahtar kelimeler: Asilidae, filogeni, yeni lokalite, COI, NADH2 Türkiye

1. Introduction

Ecological balance is known as a state of dynamic equilibrium within a community of organisms. The balance should be in the number of each species in an ecosystem. Predator groups are very important because of their stabilize feature on the specimen number in a community.

Asilidae is one of the essential groups of these predators, in controlling numerous insect groups many of which are harmful. This family represents 7104 described species and 776 genera in the world [1], 1688 species in Palearctic and nearly 241 species and 9 subspecies are reported from Anatolia [2].

There are many taxonomical and ecological studies about Turkish robber flies (e.g. [3; 4; 5]). However, despite the increasing number of phylogenetic studies about Asilidae in the world ([6; 7; 8; 9]) there is no study in Anatolia about the phylogeny of this group. Therefore, it is important to study the phylogeny of taxa belonging to Asilidae in our country.

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Promachus genus, which has 223 species worldwide, is infrequent for Turkish Asilidae fauna since five species of this genus are known from Anatolia; *Promachus canus canus* (Wiedemann, 1818), *Promachus canus leontochlaenus* Loew, 1871, *Promachus leoninus* Loew, 1848, *Promachus microlabis* Loew, 1857, *Promachus mustela* Loew, 1854 [1]. Among these the *Promachus leoninus* is distributed in Azerbaijan, Bosnia-Herzegovina, Croatia, Greece, India, Iran, Israel, Myanmar (Burma), Romania, Russia, Slovenia, Turkey on Palearctic ([10; 11; 12]). The species is known to be distributed in the western part of Anatolia on following provinces; Balıkesir, Aydın, Muğla, Ankara, Adana, Adıyaman, Afyon, Antalya, Denizli, Isparta, Mersin, Çanakkale, Eskişehir [13].

So, in the present study we aimed; (i) to provide a new data about the distribution of *P. leoninus*. (ii) to present illustration of its morphological and genital features (iii) make phylogenetic assessment with COI and NADH₂ gene regions (iv) Enter the gene bank first gene region records from Anatolia about the Asilidae.

2. Material and methods

2.1. Sampling and identification

This study is based on 21 specimens collected from 12 localities of 6 provinces in June and July on 2018 (Table 1; Figure 1). The specimens are identified with the [14] literature. Habitus and genital features were photographed with Leica DFC 490 camera, attached to a Leica MZ 16 microscope. Specimens are deposited in the Entomology collection, Department of Biology, Eskişehir Osmangazi University.

| site | Province | Locality | Date | Coordinate | | Altitude |
|------|----------|--|------------|--------------|---------------|----------|
| | | | | Latitude (N) | Longitude (E) | |
| 1 | Amasya | Yeşilöz | 26.06.2018 | 40°33'0.06" | 36°8'56.31" | 855 m |
| 2 | Erzurum | Gelinalan village | 29.06.2018 | 40° 7'59.29" | 42°33'26.62" | 1472 m |
| 3 | Erzurum | Horasan | 29.06.2018 | 40° 1'56.84" | 42° 7'41.07" | 1560 m |
| 4 | Erzurum | Mercimekli | 30.06.2018 | 40° 3'26.58" | 41°49'33.04" | 1860 m |
| 5 | Erzurum | Samikale village | 30.06.2018 | 40°23'32.36" | 41°55'58.66" | 1506 m |
| 6 | Erzurum | Oltu village | 30.06.2018 | 40°31'0.99" | 41°57'49.70" | 1322 m |
| 7 | Erzurum | Oltu village 2 | 30.06.2018 | 40°34'26.56" | 42° 1'9.94" | 1244 m |
| 8 | Erzurum | Korucuk village | 30.06.2018 | 40°30'29.18" | 41°49'53.91" | 1538 m |
| 9 | Sivas | Tödürge village | 03.07.2018 | 39°51'46.09" | 37°36'56.01" | 1338 m |
| 10 | Yozgat | Alicik village | 03.07.2018 | 39°47'8.77" | 35°58'35.92" | 1302 m |
| 11 | Bitlis | Süphan mountain/Kıskıllı village | 24.07.2018 | 38°54'55.59" | 42°54'38.71" | 2250 |
| 12 | Edirne | Aslihanlar Village | 19.06.2018 | 41°25'0.54" | 26°47'53.96" | 73 m |

Table 1. Study site information of P. leoninus specimens



Figure 1. General view of collecting site

2.2. Molecular taxonomy

In this study we studied the two gene regions of mitochondrial DNA; Cytochrome c oxidase subunit I (COI) gene region and NADH subunit 2 (NADH₂) (Table-2). Total DNA was extracted with the Thermo Scientific GeneJET Genomic DNA Purification kit. The primer sequencing was gathered from [15].

COI: 1490: 5'- GGTCAACAAATCATAAAGATATTGG-3'-3014:5'-TCCAATGCACTAATCTGCCATATTA-3'. NADH₂: N2L-J210: 5'-AATTAAGCTAATGGGTTCATACCC-3'-TW-N1284: 5' AYAGCTTTGAARGYTATTAGTTT-3'

| Table 2 | 2. Ph | ylogenetic | study | information | of specimens | |
|---------|-------|------------|-------|-------------|--------------|--|
|---------|-------|------------|-------|-------------|--------------|--|

| Locality | Province | Date | COI codes | Accession codes-COI | NADH ₂ codes | Accession codes- NADH ₂ |
|-------------------|----------|------------|--------------|------------------------|-------------------------|--|
| Yeşilöz | Amasya | 26.06.2018 | 1CamayesPrl | OL415109 | 1NamayesPrl | OL462856 |
| Gelinalan village | Erzurum | 29.06.2018 | | | 3NerzgelPrl | OL462855 |
| Gelinalan village | Erzurum | 29.06.2018 | 4CerzgelPrl | OL415106 | 4NerzgelPrl | OL462854 |
| Gelinalan village | Erzurum | 29.06.2018 | | | 5NerzgelPrl | OL462853 |
| Horasan | Erzurum | 29.06.2018 | 6CerzhorPrl | OL415105 | 6NerzhorPrl | OL462852 |
| Horasan | Erzurum | 29.06.2018 | | | 7NerzhorPrl | OL462851 |
| Mercimekli | Erzurum | 30.06.2018 | 8CerzmerPrl | OL415104 | 8NerzmerPrl | OL462850 |
| Samikale village | Erzurum | 30.06.2018 | 9CerzsamPrl | OL415103 | 9NerzsamPrl | OL462849 |
| Samikale village | Erzurum | 30.06.2018 | 10CerzsamPrl | OL415102 | 10NerzsamPrl | OL462848 |
| Oltu village | Erzurum | 30.06.2018 | 11CerzolPrl | OL415101 | 11NerzolPrl | OL462847 |
| Oltu village | Erzurum | 30.06.2018 | 12CerzolPrl | OL415100 | 12NerzolPrl | OL462846 |
| Oltu village 2 | Erzurum | 30.06.2018 | 13CerzolPrl | OL415099 | 13NerzolPrl | OL462845 |
| Oltu village 2 | Erzurum | 30.06.2018 | | | 14NerzolPrl | OL462844 |
| Küçük Korucuk | Erzurum | 30.06.2018 | 15CerzkucPrl | OL415098 | 15NerzkucPrl | OL462843 |
| Tödürge village | Sivas | 03.07.2018 | | | 16NsivtodPrl | OL462842 |

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| Alicik village | Yozgat | 03.07.2018 | 17CyozaliPrl | OL415097 | 17NyozaliPrl | OL462841 |
|-----------------------|--------|------------|--------------|----------|--------------|----------|
| Aslıhanlar village | Edirne | 19.06.2018 | 20CedasPrl | OL415096 | 20NedasPrl | OL462840 |
| Aslıhanlar village | Edirne | 19.06.2018 | 21CedasPrl | OL415095 | 21NedasPrl | OL462839 |

Polymerase chain reaction (PCR) was made in 50 μ l volume; 0,2 μ l from primers (100 pm), 1 μ l dNTP mix (10 mM), 4 μ l 1 50 mM MgCl2 (25 mM), 5 ml Standard Taq reaction Buffer 10X [containing 10 mM Tris–HCl (pH 8.3), 50 mM KCl], 0.25 TaqDNA polymerase (New England Bio labs), and 3 μ l of sample DNA. PCR cycling parameters; denaturation at 95°C for 30 sec, 35 cycles of 95°C for 20 sec., annealing at 41°C for 30 sec, elongation 72°C for 1min 40 sec. and final extension 72°C 5 min. Results were viewed with agarose gel electrophoresis by running 3 μ l of PCR product. DNA purification step and Sanger sequence analysis were carried out by Macrogen Europe (Amsterdam, the Netherlands).

2.3. Data analysis

We used 13 COI and 18 NADH₂ sequences of *Promachus leoninus* for analyses. Two outgroups are used for COI dataset; *Leptogaster cylindrica* (Accession code: OL415107) and *Dysmachus bifurcus* (Accession code: OL415108) (our own samples) and one outgroup is gathered from NCBI for NADH₂ dataset (Accession code: EF216223.1) *Drosophila guanche*. All of the sequences were checked manually with SEQUENCER v. 4.1 (Gene Codes Corporation). The alignment was made with MEGA v.7 [15] and the program was used for calculating the conservative, variable and parsimony-informative sites. Haplotypes and their frequency were determined with DnaSP v.5 [17]. Sequences are deposited in the Genbank database (accession number can be seen at Table 2). The best fit evolutionary model for the data matrix was estimated by jModelTest v.0.1.1 [18].

Sequences were analyzed with three different phylogenetic methods; (i) Maximum parsimony (MP) by PAUP Version 4.0b10 [19], with 100 random additions, with nearest neighbor interchange (NNI) algorithm and heuristic search approach. (ii) Maximum Likelihood (ML) with raxmlGUIversion 1.5 [20] with ML-rapid 1000 bootstrap option. (iii) Bayesian phylogenetic inference (BI) analyses with MrBayes v.3.1.2 [21] program, four simulations of Markov chains, 10 M generations and sampling every 100th generations and with the program the 1000 trees were discarded as burn-in.

We applied to the COI matrix two different analyses to test species limits. DNA sequence-based analysis TCS [22] and Automatic Barcode Gap Discovery (ABGD) programs were applied to the dataset to make species delimitation test.

3. Results

3.1. Plot summary of morphology

Body; slender with a dense hair, the color of the body is dark metallic which is bright to pale, with yellow to orange bands, with these colors the species reminds bees. Thorax; pronotum with hairs. Wings are transparent and clear (Figure 2b), legs are strong and also covered with striped with black and yellow dense hair (Figure 2a). Head; typically slender with big black dichoptic eyes (varies to green blue), antenna's 3rd segment is elongated thinning at the end (Figure 2c). Mouthparts are black and very strong. The aedeagus is conspicuously long and can be seen at the end of the abdomen (Figure 2d-f). Gonocoxites are not fused. Dististylus broad, curved with a pointed end and with setae at the ventral margin. Hypandrium short, with rounded sides and laterally short setae.



Figure 2. a-i. A male *Promachus leoninus* Loew 1848. a-lateral view of specimen b-wing c-dorsal sight of the head with antennae d-last abdominal segment with aedeagus e-epandrium f-lateral view of aedeagus g-gonocoxite and dististylus h-hypandrium i- proctiger

3.2. Molecular assessment with mtDNA gene regions

The sequences of *Promachus leoninus* (13 COI and 18 NADH₂) were checked with BLAST program after alignment. Data set for COI with 15 sequences (with two outgroups) and 1222 base pairs and for $NADH_2$ with 19 sequences (with one outgroup) and 1001 base pairs were obtained. The tables can be checked for detailed information (Table-3 about data sets and Table-4 for haplotypes for two gene regions).

| | Number of Sequence | Total Number of Sites | Conserved | Variable | Pars- inf | Number of Haplotypes | Haplotype Diversity |
|-------------------|-----------------------|-----------------------------|-----------|----------|--------------|-------------------------|------------------------|
| COI | 15 | 1222 | 901 | 321 | 117 | 13 | 0,9810 |
| NADH ₂ | 19 | 1001 | 715 | 286 | 31 | 10 | 0,7836 |

Table 3. Sequence and haplotype informations of COI and NADH₂ gene regions

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| COI gene region | 1 71 | NADH ₂ gene region | | |
|---------------------|--------------|-------------------------------|-------------------------|--|
| Number of | h: 12 | Number of healetings | h:10 | |
| haplotypes | 11. 15 | Number of napiotypes | | |
| Haplotype diversity | Hd: 0,9810 | Haplotype diversity | Hd: 0,7836 | |
| Hap_1: 1 | 21CedasPrl | Hap_1: 1 | 21NedasPrl | |
| Hap_2: 1 | 20CedasPrl | Hap_2: 1 | 20NedasPrl | |
| Hap_3: 1 | 17CyozaliPrl | Hap_3: 1 | 17NyozaliPrl | |
| Hap_4: 2 | 15CerzkucPrl | Hap_4: 1 | 16NeintodDrl | |
| | 13CerzolPrl | | TOINSIVIOUFII | |
| | | | 15NerzkucPrl | |
| | | | 13NerzolPrl 9NerzsamPrl | |
| Hap_5: 1 | 12CerzolPrl | Hap_5: 9 | 8NerzmerPrl 7NerzhorPrl | |
| | | | 6NerzhorPrl 5NerzgelPrl | |
| | | | 4NerzgelPrl 3NerzgelPrl | |
| Hap_6: 1 | 11CerzolPrl | Hap_6: 2 | 14NerzolPrl 11NerzolPrl | |
| Hap_7: 1 | 10CerzsamPrl | Hap_7: 1 | 12NerzolPrl | |
| U.m. 9, 2 | 9CerzsamPrl | II.m. 9, 1 | EF216223.1_Drosophil | |
| пар_о. 2 | 6CerzhorPrl | пар_8. 1 | Outgroup | |
| Hap_9: 1 | 8CerzmerPrl | Hap_9: 1 | 10NerzsamPrl | |
| Hap_10: 1 | 4CerzgelPrl | Hap_10: 1 | 1NamayesPrl | |
| Hop. 11, 1 | 11ckirder | | | |
| пар_11. 1 | Outgroup | | | |
| Hop. 12: 1 | 1dbkirkof | | | |
| пар_12: 1 | Outgroup | | | |
| Hap_13: 1 | 1CamayesPrl | | | |

Table 4. Detailed information about Haplotypes

3.3. COI gene region results

The dataset file was converted into different formats with Mesquite and Alter programs for use in phylogenetic analysis. Haplotype diversity was calculated as 0, 9810 for COI with DnaSP v.5. There is no haplotype sharing; all the haplotypes are unique sequences except haplotypes 4 and 8 (Table-4). The jModelTest v.0.1.1 suggested the evolutionary model as GTR+G according to AIC (Akaike Information Criterion), with p-inv = 0.2571 gamma shape = 0.4450.

Maximum likelihood, Maximum parsimony, Bayesian Phylogenetic inference analyses are result in a resembling tree topology, therefore all results are shown on BI tree (Figure 3). Haplotypes of individuals were grouped (with strong branch node values) according to population structure as expected. The Edirne, Erzurum and Yozgat-Amasya populations are branched as ingroups. The network analysis is also consistent with the phylogenetic analysis results (Figure 4). We can also observe same the population ingroups in this analysis. TCS (%90) Species delimitation test suggested two different species while TCS (%95) test suggested three populations as three different species, but morphological differences were not observed among the individuals of the population. Also, ABGD test is suggesting that all of the haplotypes belong to single species. Therefore, these populations can be considered as diverging populations and not as separate species.



Figure 3. ML, MP, BI analyses results of COI gene region. Bootstrap values (MP-ML) and posterior probability values (BI) are shown. Respectively; Black is for BI, red is for MP and blue is for ML. (*) indicates 50% and below values or not supported by the respective analyses. ABGD and TCS Species delimitation tests are mapped on the tree.



Figure 4. Network analysis results of COI gene region

3.4. NADH₂ gene region results

Haplotype diversity was calculated as 0, 7836 for NADH₂ with DnaSP v.5. There is no haplotype sharing; all the haplotypes are unique sequences except haplotype 8 (Table-4). The jModelTest v.0.1.1 suggested the evolutionary model as GTR+G according to AIC (Akaike Information Criterion), with p-inv = 0.5140 gamma shape = 100.0000.

All the analyses (ML, MP and BI) are resulted in the same tree topology, shown on MP tree (Figure 5). As seen in the phylogenetic analyses made with the COI gene region, the haplotypes are branched according to the populations, but the branch support values are lower. The branch support value (BI: >50, ML: 54, MP: 70) at the branch node of the Edirne and Yozgat-Amasya populations seems to be quite low (Figure 5). We can observe the same result on network analysis. Network analyses resulted in same way as in phylogenetic analyses. We can see the haplotypes are grouped in three different populations (Yozgat-Amasya population, Edirne population and Erzurum population) (Figure 6).

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TCS (%95) Species delimitation test suggested two different species but morphological differences were not observed among the individuals of the population. All of the haplotypes are belong to single species according to TCS (%90) and ABGD tests.



Figure 5. ML, MP, BI analyses results of NADH₂ gene region. Bootstrap values (MP-ML) and posterior probability values (BI) are shown. Respectively; Black is for BI, red is for MP and blue is for ML. (*) indicates 50% and below values or not supported by the respective analyses. ABGD and TCS Species delimitation tests are mapped on the tree.



Figure 6. Network analysis results of NADH₂ gene region

4. Conclusions and discussions

The distributional area of *P. leoninus* was known from South and West part of Anatolia. 21 specimens (14 male and 7 female) are collected from east part of Anatolia, Middle Black Sea and the Trace region (Figure 7). The specimens examined here present a new distribution record for *P. leoninus*, which is considerably enlarging the distributional area to east part of Anatolia and the Trace region.

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Figure 7. Previous and new locality records of Promachus leoninus

Therefore, there is known from our records that these species are active at 10:00 am in the morning to 8:00 pm on the sunset. The optimal temperature for their activity is between 21- 37 $^{\circ}$ C degrees, based on our observations. Further studies are needed to do for determining the host preference of *P. leoninus*.

This is the first study conducted in our country on the phylogeny of this group. The new distribution regions of *P. leoninus* are grouped into 3 different populations. Phylogenetic analyses were carried out with the COI and NADH₂ gene regions. The analyses performed similar results in both gene regions. Haplotypes belonging to the same populations branched together in the phylogenetic trees. While this branching in the COI gene region is supported by high branch support values, it has been observed that these values are lower in NADH₂.

Similar results were obtained in the haplotype Network analysis as in the phylogenetic analyses for both gene regions. In some of the species delimitation tests; populations were proposed as different species. However, individuals belonging to these populations were morphologically examined once again and it was seen that all individuals share the same morphological characters, herewith we can use the term "diverging populations" instead of "different species".

In order to reveal the systematic and phylogenetic status of the species more clearly, a broader evaluation with the Balkan and Caucasian populations is required. More gene regions or next generation sequencing methods can be studied.

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